

Studying antioxidant, anti-inflammatory activities and antimicrobial of Iron nanoparticles biosynthesized from water extract of *Mentha pulegium L*

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Abstract:

Biosynthesis of Iron nanoparticles from *Mentha pulegium L* by simple method by using water extract of *Mentha pulegium L*. The iron nanoparticles characterized by UV-visible spectroscopy, (FTIR) Fourier transform infrared spectroscopy, XRD and Scanning electron microscopy (SEM). The biological activity evaluation by anti-inflammatory and anti oxidant using evaluated in vitro 2,2-diphenyl-1-picrylhydrazyl (DPPH) assay, ABTS radical scavenging assay Iron chelating activity assay . The results also revealed that the water extract of *Mentha pulegium L* and its iron nanoparticles represented the most potent in vitro antioxidant effect. Iron nanoparticles inhibited the growth of bacterial and fungal, these results suggested that iron nano particle could be an interesting to control pathogenic micro organisms.

Keywords: Iron nanoparticles, plant extracts, *Mentha pulegium L*, antioxidant activity, antiinflammatory activity.

Introduction:

Now a days great efforts were made to use green and environmentally friendly methods for synthesis nano materials, these efforts involve the use of plant or fruit extracts as detergent (1,2). Metal ions reduce by plant extracts in a shorter time in compared with microbes, depending upon plant type and concentration of phytochemicals(3). Nanoparticles are synthesized within a few minutes or hours, whereas microorganism-based methods require a longer time (4, 5). Our work describes a green and rapid method using *Mentha* leaves, water extract solution for the biosynthesis of iron oxide nanoparticles without any additives protecting nanoparticles from aggregating. The current green method using rapid precursors of *Mentha* leaves extract provides high-yield nano materials with a good optical properties, and the method can be used to prepare nano oxides of other interesting materials. The Fe_3O_4 -

NPs were prepared by using ferric sulfate as iron precursor and *Mentha* leaves extract as reducing agent and stabilizer.

Materials and Methods

Fe₃O₄-NPs were synthesized using the leaves extract of *Mentha*. Five gram powdered sample from *Mentha pulegium L* extracted with 150 ml D. W at room temperature. 0.1M FeSO₄ (40 ml) of water extract was added to 40 ml leaves extract drop wise with continuous still changing the color of solution from pale yellow to black . The solution was centrifuged at 10,000 rpm, at 4°C for 15 min and pellet was dissolved in distilled water for periodic probe sonication for 5 min at 30 ±0.5°C. Nano suspension thus obtained was dried in oven.

Characterization of Iron Nanoparticle

The structure and purity of Fe₃O₄-NPs identified by X-ray diffraction measurement. Recorded in the range 500–4000 nm. UV-Vis spectral analysis was done by using UV-Vis, at the range of 200 -600 nm and observed the absorption peaks at nm regions due to the excitation of surface Plasmon vibrations in the Fe₃O₄-NPs solution, which are identical to the characteristics UV-visible spectrum of metallic Iron and it was recorded. Scanning Electron Microscope (SEM) to characterize mean particle size and morphology of Iron oxide nanoparticles, SEM was performed using Shimadzu SEM machine of 20 KV of accelerating voltage. FT-IR spectra of the Fe₃O₄-NPs were recorded in the range 500–4000 nm by (Shimadzu, Tokyo, Japan).

DPPH radical scavenging assay:

To Fe₃O₄-NPs solution of DPPH (1mM) an equal volume of the nano particle solution dissolved in water added at various concentrations from 100 to 750 µg/ml in a final volume of 1.0 ml. An equal amount of alcohol was added to the control. After 20 min. absorbance was measured at 517 nm. Experiment was performed in triplicate (6).

ABTS radical scavenging assay:

To the reaction mixture containing 0.3 ml of ABTS radical, 1.7 ml phosphate buffer and 0.5 ml Nano particle was added at various concentrations from 2 to 500 µg/ml. Blank was carried out without drug. Absorbance was recorded at 734 nm. Experiment was performed in triplicate (7).

Iron chelating activity assay:

The chelation of ferrous ions by iron nanoparticles was estimated by method(8). Take 50 μ l of FeCl_2 (2 mM) was added to 1 ml of different concentrations of the extract (0.2, 0.4, 0.8, 1.6 and 3.2 mg/ml). The initiation of reaction by addition (0.2) ml of 5 mM ferrozine solution. The mixture was vigorously shaken and left to stand at room temperature for 10 min. The absorbance of the solution was thereafter measured at 562 nm. The percentage inhibition of ferrozine-Fe $^{2+}$ complex formation was calculated as $[(A_0 - A_s) / A_s] \times 100$, where A_0 was the absorbance of the control, and A_s was the absorbance of the extract/ standard. Na_2EDTA was used as positive control.

Denaturation inhibition of albumin

One percent(1%) Human albumin was incubated at 37 C° for 20 minute, heated at 51 °C cooling. Measured the turbidity at 660 nm by UV Visible Spectrophotometer. The Percentage of denaturation inhibition calculated by this equation = $(\text{Abs Control} - \text{Abs Sample}) \times 100 / \text{Abs control}$.

Detection membrane RBCs stabilization

Human blood cells centrifuged by 3000 rpm for 10 min, precipitate washed with normal saline then suspended to 10% v/v by normal saline, suspension incubated at 56°C in water bath for 30 min, centrifuged at 3000 rpm for five minute detection the absorbance at 560 nm. The Percentage of haemolysis inhibition calculated (9):

$$\% \text{ inhibition} = (\text{Abs control} - \text{Abs sample}) \times 100 / \text{Abs control}$$

Microorganisms

Culture of, *Pseudomonas aeruginosa*, *Bacillus subtilis*, *Escherichia coli*, and *Klebsiella pneumoniae* species of bacteria and *Aspergillus niger* and *Aspergillus flavus* *Candida spp.* From public health center.

Antibacterial activity

The antibacterial activities of iron nanoparticle were carried out by disc diffusion method(9). Nutrient agar medium plates were prepared, sterilized and solidified. After solidification bacterial cultures were swabbed on these plates. Using filter disc immersed in iron nanoparticles solution (25 mg/ml) and placed in the nutrient agar plate and kept for incubation at 37°C for 48 hours. Zones of inhibition for control. The experiments were (6).

Antifungal activity

The fungal suspension of *Aspergillus flavus* *Candida spp* was prepared in normal saline by transferring the organism from fresh cultures (1×10^8 cells/ml). To determine the antifungal cultures were poured with a Sabourauds glucose agar medium (30 ml) in petri plates (90 mm). Sterilized filter paper discs (Whatman No. 1; 6 mm in diameter) soaked in 25 mg/ml concentrations of iron nanoparticle were taken out with sterilized forceps and air-dried and placed on plates with the different organisms. The plates were incubated for 7 days at 25°C. After incubation, the inoculated plates were observed for inhibition zone(7).

of *Mentha pulegium L* from pale yellow to black color Figure (1). The UV- VS spectrum of Fe₃O₄-NPs of the *Mentha pulegium L* extract peaks at 400-475 nm indicate the formation of iron nanoparticles Figure (2).



Fig (1) Biosynthesis of nanoparticle

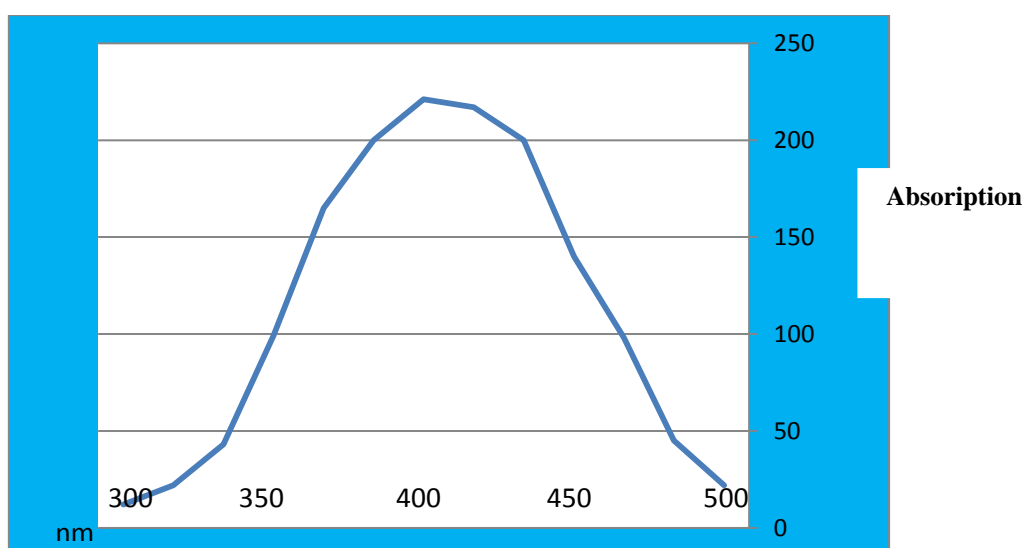


Fig (2) Shows absorbance of samples by different wave length.

SEM analysis

The image of SEM for iron nanoparticles were synthesized from *Mentha* water. It was shown that relatively spherical nanoparticles were formed with diameter of 15 to 25.6 nm. The nanoparticles were not in direct contact even within the aggregates, indicating stabilization of the nanoparticles by a capping agent (9, 10) (Fig 3).

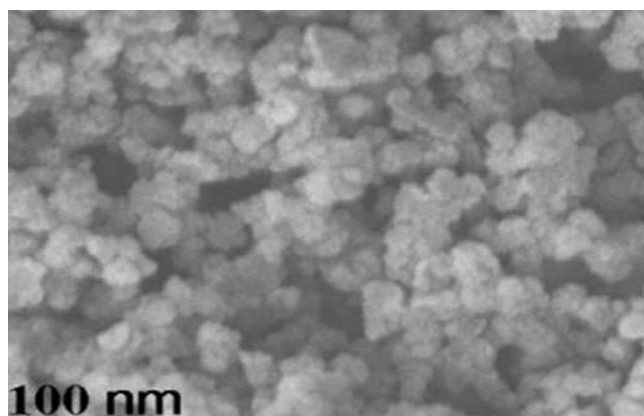


Figure (3) Showed iron nanoparticles by SEM.

Determination the *Mentha pulegium L* extract functional groups and detection their role in the synthesis of Iron nanoparticles by FTIR analysis bands in different regions of the spectrum for Iron NPs, respectively were analyzed and are shown in Figure(4). The absorption of peak at around 3,394 cm^{-1} returns to the O–H stretching vibrations of phenols and carboxylic acids. The peak located at around 2,355 cm^{-1} was attributed to the N–H stretching or the C = O stretching vibrations. The peak located at 1,641 cm^{-1} could be assigned to the C = O stretching in carboxyl or C = N bending in theamide group,The peak at 3,388 cm^{-1} may indicate the involvement of O–H functional group in the synthesis of nanoparticles. Peaks (from 1641 to 1643 cm^{-1}) indicated carboxyl or amino groups in synthesis nanoparticle. The peak at 771 cm^{-1} and 760 cm^{-1} corresponds to C–H stretching of aromatic compounds. The formation of Fe_3O_4 was characterized by two absorption bands at 535 and 307 cm^{-1} which correspond to the Fe–O bond in magnetite (11) From the FTIR result, the soluble elements present in *Mentha pulegium L* extract acted as capping agents preventing the aggregation of nanoparticles in solution, and thus playing a relevant role in their extracellular synthesis (12).

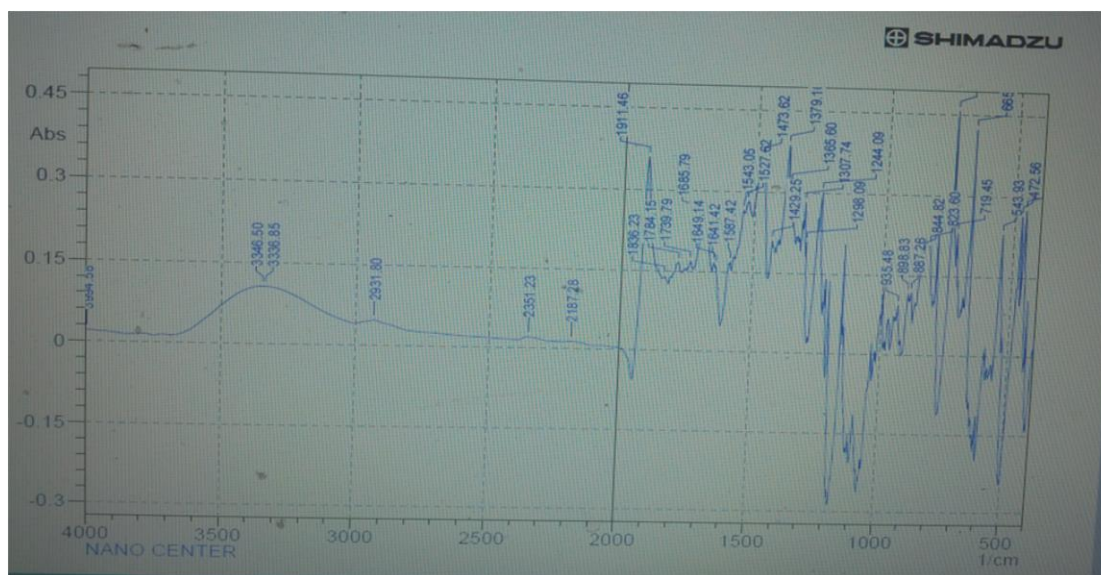


Figure 4. A FT-IR spectrum for the Fe₃O₄-NPs.

Using different models for detection antioxidant activity. It was observed that free radicals scavenged by the test compounds in a concentration dependent manner up to the given concentration in all the models. The percentage of scavenging and IC₅₀ values were calculated for all models given in Table (1). The anti oxidative mechanism resulted from metal chelation, free radical scavenging (hydrogen-donation free radical quenching, or co-operative effects of these properties (12). The ability of nano particles to donate hydrogen was checked by using stable free radical DPPH was formed from the scavenger the reaction is monitored by the decrease of the absorbance at 518nm (13).

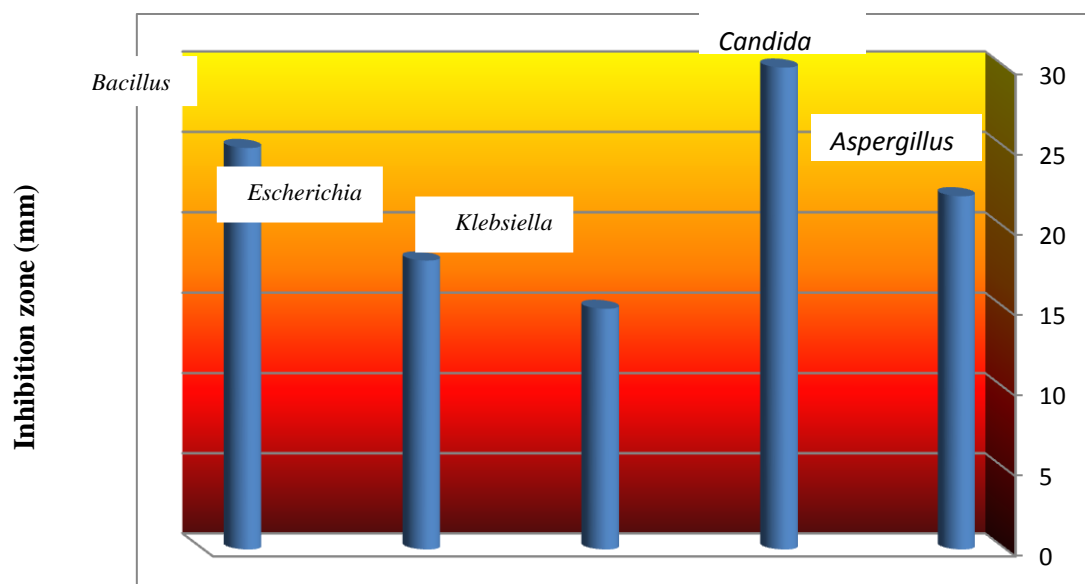
Table (1) : Comparison of IC₅₀ values of Iron nanoparticle in comparison with standard.

No	Test	IC ₅₀ µg / ml
1	DPPH radical scavenging activity	22.5
2	ABTS radical scavenging activity	33.4
3	Iron chelating method	52.8

Antimicrobial and Antifungi

Fig. 5. Shows the antimicrobial activity of synthesized Iron nanoparticles against five different bacteria and fungi such as *Bacillus subtilis*, *Escherichia coli*, *Proteus vulgaris* and *Klebsiella pneumoneae*, *Candida spp* and *Aspergillus flavus*. As it showed a clear inhibition zone, the synthesized nanoparticles were highly effective in their activity against

Candida spp and *Bacillus subtilis* folloed by *Aspergillus flavus*, *Eecherichia coli* and *Klebsiella pneumoneae*. The Iron nanoparticles synthesized via green route are highly toxic towards fungal species also when compared to bacterial species. our findings of suggested that the inhibition of oxidation based biological process by penetration of metallic nano sized particles across the microsomal membrane (13),(14).



Fig(5) Showed inhibition% activity of Iron nano particle on microorganism

Inhibition albumin denaturation

Table (2) showed maximum inhibition of albumin denaturation was 77.5% observed at 450 µg/ml of nano particle in compared with aspirin at the same concentration. This result investigated that nano particle under study was effective in inhibiting albumin denaturation. Denaturation of proteins refers to cause of inflammation (15). So for this we were studying protein denaturation to evaluate anti-inflammatory activity of iron nanoparticles.

Table (2) showed the results of albumin denaturation Inhibition.

Concentration (µg/ml)	Absorbance (nm)	inhibition% of hemolysis
<u>CONTROL</u>	<u>0.57±0.01</u>	=
<u>150</u>	<u>0.32± 0.014</u>	<u>37</u>
<u>250</u>	<u>0.29± 0.04</u>	<u>59.6</u>
<u>350</u>	<u>0.15± 0.02</u>	<u>67.8</u>
<u>450</u>	<u>0.09± 0.07</u>	<u>77.5</u>
<u>450 ASPIRIN</u>	<u>0.15±0.01</u>	<u>81.7</u>

Haemolysis Heat Induced Haemolysis

Table (3) showed the results activity of inhibiting haemolysis at different concentrations of iron nanoparticles. The effective concentration was 300µg/ml in compare with standard drug Declofenac 400µg/ml gave agood protection against damaging effect of heat solution.

Table (3) Showed the inhibition of protein denaturation.

Concentration (µg/ml)	Absorbance (nm)	inhibition% of hemolysis
<u>CONTROL-</u>	<u>0.52±0.01</u>	<u>-</u>
<u>100</u>	<u>0.34±0.03</u>	<u>30</u>
<u>200</u>	<u>0.22±0.03</u>	<u>40.2</u>
<u>300</u>	<u>0.10±0.04</u>	<u>65.6</u>
<u>400</u>	<u>0.15±0.03</u>	<u>80.6</u>
<u>400</u> Declofenac	<u>0.13±0.07</u>	<u>85</u>

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